



Review

405 nm light technology for the inactivation of pathogens and its potential role for environmental disinfection and infection control

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SUMMARY

Background: Although the germicidal properties of ultraviolet (UV) light have long been known, it is only comparatively recently that the antimicrobial properties of visible violet–blue 405 nm light have been discovered and used for environmental disinfection and infection control applications.

Aim: To review the antimicrobial properties of 405 nm light and to describe its application as an environmental decontamination technology with particular reference to disinfection of the hospital environment.

Methods: Extensive literature searches for relevant scientific papers and reports.

Findings: A large body of scientific evidence is now available that provides underpinning knowledge of the 405 nm light-induced photodynamic inactivation process involved in the destruction of a wide range of prokaryotic and eukaryotic microbial species, including resistant forms such as bacterial and fungal spores. For practical application, a high-intensity narrow-spectrum light environmental disinfection system (HINS-light EDS) has been developed and tested in hospital isolation rooms. The trial results have demonstrated that this 405 nm light system can provide continuous disinfection of air and exposed surfaces in occupied areas of the hospital, thereby substantially enhancing standard cleaning and infection control procedures.

Conclusion: Violet–blue light, particularly 405 nm light, has significant antimicrobial properties against a wide range of bacterial and fungal pathogens and, although germicidal efficacy is lower than UV light, this limitation is offset by its facility for safe, continuous use in occupied environments. Promising results on disinfection efficacy have been obtained in hospital trials but the full impact of this technology on reduction of healthcare-associated infection has yet to be determined.

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Introduction

Although intensive efforts over recent years are making an impact, healthcare-associated infections (HCAIs) still regularly occur and continue to pose a major challenge. In addition to the significant morbidity and financial costs, concern over

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contraction of HCAs is one of the greatest fears of patients being admitted to hospital.¹ Infection control procedures such as handwashing are of critical importance in addressing the HCAI problem; however, greater awareness of the hospital environment as a source of nosocomial pathogens has led to renewed focus on hospital cleaning and disinfection. Whereas effective physical cleaning remains essential for infection control and aesthetic reasons, there has been an upsurge of interest in the development of new cleaning and decontamination technologies.^{2,3} Several of these employ novel methods of delivering antimicrobial chemicals, whereas others use the antimicrobial properties of light to enhance disinfection, and it is this latter approach that forms the topic of this review.^{4–6}

The most germicidal wavelengths of light fall within the ultraviolet (UV) range and UVC (240–260 nm) irradiation has traditionally been used for disinfection, particularly for air and medical device decontamination applications.^{7–9} More recently the antimicrobial properties of violet–blue visible light have emerged as an area of increasing research interest. Although less germicidal than UVC light, violet–blue light with wavelengths in the region of 405 nm has proved effective for inactivation of a range of microbial species, and exploitation of these wavelengths may provide alternative methods of antimicrobial treatment for infection control applications. This paper supplies a brief background on the use of light for environmental decontamination applications within hospitals before presenting a detailed description of the broad spectrum antimicrobial effects of violet–blue light and how this knowledge has led to the development and clinical evaluation of a 405 nm light environmental disinfection system. In addition to environmental decontamination applications, other potential uses of violet–blue light for infection control purposes such as skin and wound treatment have been highlighted in recent literature but these topics are out with the scope of the current review.^{10–17}

Inactivation of micro-organisms by light in the hospital environment

Records of observations on the antibacterial effects of light go back to the latter part of the 19th century and these early historical observations have been documented by Kowalski.¹⁸ The germicidal effects of light received further attention during the early part of the 20th century and the appreciation of the decontamination effect of light was translated into early hospital design features where natural ventilation and exposure to sunlight were regarded as beneficial.¹⁹ The roles of sunlight and natural ventilation for controlling the transmission of infections within healthcare settings has recently been reviewed by Hobday and Dancer, who provide a detailed record of the early–mid-20th century observations on the effects of natural sunlight on a wide range of nosocomial pathogens.²⁰ Although natural light and ventilation were originally considered beneficial, modern hospital design has tended to reduce these features. Recent interest in the application of ‘artificial’ lighting within hospitals has been with regard to energy reduction issues but also how lighting can affect the mood and circadian rhythm of patients.^{21,22} Light from artificial sources with wavelength emission in the UV range can have significant antimicrobial effects and new technologies for hospital decontamination have been developed around this concept.^{6,23–25}

The most widespread applications of ultraviolet germicidal irradiation (UVGI) has been for air and water disinfection, as well as for decontamination of devices.^{26–28} More recently, with the increased emphasis that has been directed towards enhanced decontamination of the hospital environment, novel technologies have been developed for the rapid delivery of UVC radiation to exposed surfaces in clinical areas. Several of these are automated or manually positioned robotic systems using either continuous or pulsed UV emission sources.^{6,25} Detailed information on UVGI and other ‘no-touch’ automated room disinfection systems is provided in a recent review by Otter et al.⁶

Antimicrobial effects of violet–blue light

Until relatively recently light within the visible spectrum (400–700 nm) was considered to have little biocidal effect compared to UVC light due to the lower photon energy of these wavelengths. Wavelengths of violet–blue light, particularly around 405 nm, have, however, been shown to possess antimicrobial capabilities, and there is scope for exploiting these wavelengths for the control of problematic micro-organisms in many areas of application including the disinfection of air and exposed surfaces in the clinical environment. The following section provides an overview of the antimicrobial inactivation mechanism, and the antimicrobial efficacy of high-intensity 405 nm violet–blue light.

Violet–blue light inactivation mechanism

Investigations into the mechanism of action of 405 nm violet–blue light indicate that **photodynamic inactivation occurs as a result of the photo-excitation of intracellular porphyrin molecules within the exposed bacterial cells.** Laboratory studies have shown that a range of violet–blue light wavelengths in the region 400–425 nm can be used for bacterial inactivation; however, optimal antimicrobial activity has been found at 405 nm.^{29–35} This peak in activity correlates with the absorption maximum of porphyrin molecules, termed the Soret band, being in this wavelength region.³⁶ Exposure to light of this wavelength induces an oxygen-dependent photo-excitation reaction within exposed micro-organisms, where **excited porphyrins react with oxygen or cell components to produce reactive oxygen species (ROS), causing oxidative damage and microbial cell death.**^{29,37–41} Cell death has been accredited to **oxidative damage to the cell membrane, with a recent study demonstrating disruption of the cytoplasmic content and cell walls of exposed *Staphylococcus aureus*, and it is likely that, due to the non-selective nature of ROS, multi-target damage will be induced in the microbial cells.**¹⁰

Antimicrobial effects of violet–blue light

Extensive laboratory studies have shown that 405 nm light, and the wider violet–blue light wavelengths, have a broad spectrum of activity, with successful inactivation demonstrated for a wide range of organisms, including antibiotic-resistant bacterial strains such as methicillin-resistant *Staphylococcus aureus* (MRSA).^{30–32} Bacterial species which have demonstrated susceptibility include HCAI-associated organisms, including *S. aureus*, *Clostridium difficile*, *Acinetobacter*

baumannii, *Escherichia coli*, *Staphylococcus epidermidis*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Streptococcus pyogenes* and *Mycobacterium* spp.^{29–33,42,43} Bacterial sensitivity to violet–blue light inactivation tends to be species dependent; however, the general trend suggests that **Gram-positive bacteria tend to be more susceptible to inactivation than Gram-negative species.**^{32,44}

Two of the most significant pathogens associated with HCAI are MRSA and *C. difficile*, and vegetative cells of these species both show susceptibility to violet–blue light inactivation. Vegetative cells of *C. difficile* are particularly sensitive to inactivation, and this is likely to be due to this organism being an obligate anaerobe, giving it increased sensitivity to oxidative damage.³³ *C. difficile* spores are a significant issue for infection control, particularly due to their prolonged survival in the environment, and their resilience to disinfection technologies is well documented.^{45–47} ***C. difficile* spores can be successfully inactivated by exposure to 405 nm light, but, as expected, significantly higher doses (~50 times) are required for inactivation compared to vegetative cells.**³³

Laboratory studies have demonstrated the successful antimicrobial efficacy of violet–blue light for the inactivation of bacterial contamination in liquid, **artificially seeded on surfaces, and most recently in biofilms.**^{10,11,29–32,34,42,44,48} Within the clinical environment, biofilm formation is a major cross-contamination risk, with the presence of patient fluids such as saliva, blood and urine influencing biofilm adhesion and development on surfaces.⁴⁹ Indeed, a recent study attributed the presence of *Pseudomonas aeruginosa* biofilms on sinks to the acquisition of infections, with a 33% death rate.⁵⁰

Although the germicidal efficacy of blue light is lower than that of UV light – UV inactivation typically required doses of the order of milli-joules rather than joules, as is the case with violet–blue light – significant bacterial inactivation can still be demonstrated, with up to 9-log₁₀ orders of reduction being achieved by Maclean et al.^{32,51,52} A major advantage of violet–blue light inactivation is that the **susceptibility of strains isolated from the clinical environment is similar to their laboratory type strain counterparts, i.e. clinical isolates do not show enhanced resistance and thus can be inactivated by 405 nm light with no inherent problems.**³² Also, it has recently been demonstrated that sublethally damaged bacterial cells are more susceptible to light inactivation; therefore, there is great potential for bacterial contamination that has been sublethally stressed by desiccation and disinfectants during routine cleaning of the hospital environment to be more susceptible to inactivation by exposure to violet–blue light.⁴⁸

In addition to clinically relevant bacteria, the effectiveness of 405 nm light for microbial inactivation has also been demonstrated against bacterial species associated with food-borne infection including *Listeria*, *Campylobacter*, *Shigella* and *Salmonella* spp.; pathogens *Helicobacter pylori*, *Chlamydia* and *Propionibacterium acnes*; oral periodontal pathogens; and **fungal organisms including moulds and yeasts such as *Candida*.**^{5,29,32,34,37,43,53–56} To date, the effect of violet–blue light on viruses has not been fully determined; however, it is expected that, due to the hypothesized involvement of porphyrins in the inactivation mechanism, it is unlikely that viruses will be highly susceptible to light exposure alone, and may require the addition of photosensitizing material to enhance viricidal activity.⁵⁷

Use of 405 nm violet–blue light for hospital disinfection

The wide antimicrobial spectrum of activity combined with the ability to apply light intensities safe for human exposure make violet–blue light ideal for decontamination of occupied environments, and the development of a system which uses high-intensity narrow spectrum (HINS) 405 nm light for environmental disinfection of the clinical environment has been recently described.^{58–60} This new disinfection technology, termed the HINS-light environmental decontamination system (EDS), is a ceiling-mounted lighting system designed for the reduction of environmental contamination in hospital wards and other areas of the healthcare environment. The antimicrobial light from the system is generated from a matrix of light-emitting diodes (LEDs) which emit low-irradiance violet–blue light with a narrow spectral profile centred on 405 nm.⁵⁸ **The output of the antimicrobial light has been set to ensure, with reference to international guidelines, that the light source does not pose a blue light hazard and is safe for use in occupied environments.**^{61,62} Although biocidal, the 405 nm wavelengths are well below the blue light wavelengths which can impact on human health, particularly in the region of 440 nm which is associated with photoretinitis, and 480 nm which influences mood and circadian rhythm in humans (Figure 1). It is interesting also to note that, when comparing the susceptibility of mammalian cells and bacteria to 405 nm light, **mammalian keratinocytes and osteoblasts were considerably more resistant and could be exposed to bactericidal levels of 405 nm light with no loss of cell viability.**^{10,11,63} The increased resistance of mammalian cells is likely due to the fact that these cells have much more advanced mechanisms for coping with oxidative damage compared to the more primitive microbial cells.

For practical application as an overhead light source, incorporation of white LEDs into the HINS-light EDS system ensures that the illumination output is predominantly white, thus blending with the standard room lighting.⁵⁸ The system is designed to be operated continuously, providing ongoing disinfection of the air and all exposed environmental surfaces within the treated area, with no disruption to day-to-day hospital procedures or patient care. Laboratory testing of the system confirms the efficacy for inactivation of a range of bacterial pathogens associated with HCAI.⁶⁴ As mentioned, the low irradiance levels employed by the system were deliberately selected to enable continuous disinfection in occupied environments, and therefore require sufficient time to exert the antimicrobial effect. **Significant inactivation of microbial contamination on simulated laboratory surfaces can be achieved by ~1–2 h light exposure; however, inactivation kinetics are likely to be significantly enhanced in the 'real' clinical environment due to the stressed and desiccated state of the micro-organisms.**^{48,64}

Clinical assessment of 405 nm light for environmental disinfection

Several published studies have presented results from clinical assessment of this 405 nm light system for continuous environmental decontamination of single-bed isolation rooms.^{58–60} Evaluation of the technology has been carried out in isolation rooms within two main clinical areas: a burns unit and an intensive care unit (ICU).

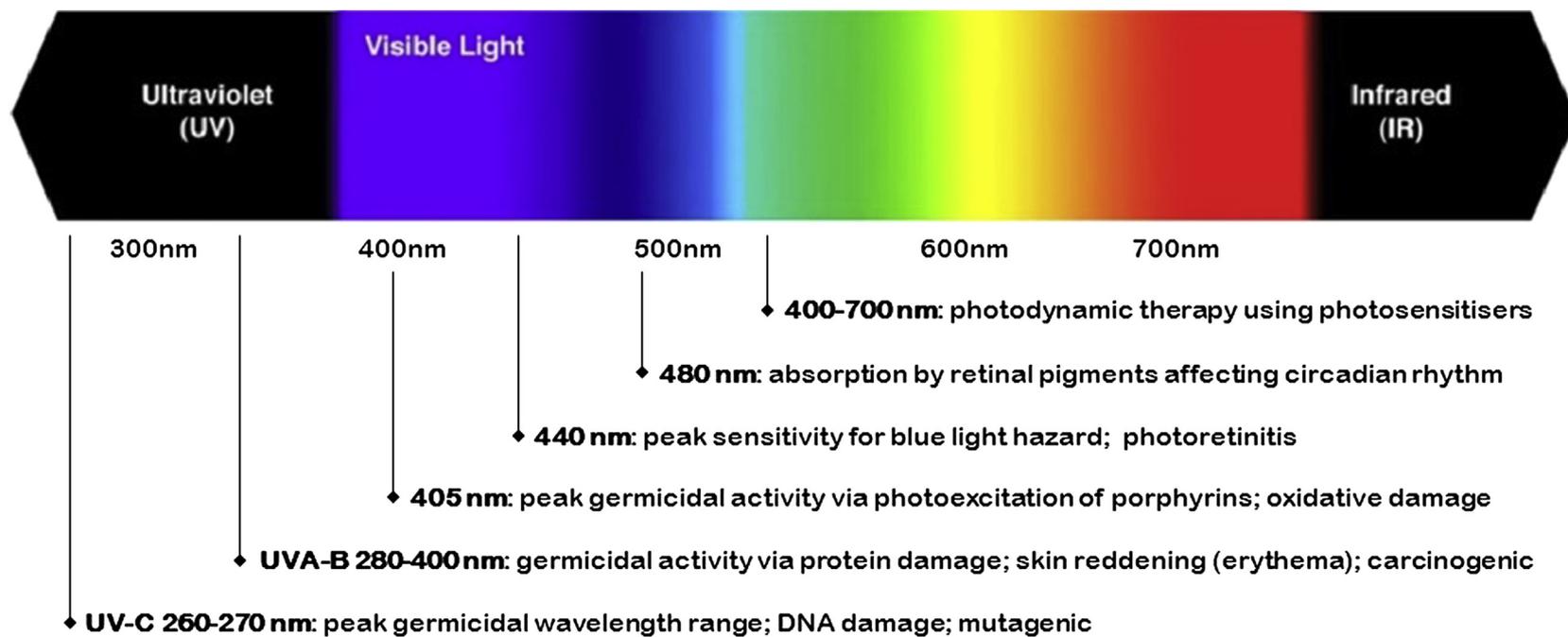


Figure 1. Ultraviolet (UV), visible light and infrared regions of the electromagnetic spectrum. Highlighted are key UV and violet/blue wavelengths with details of their germicidal action and safety aspects. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

For evaluation, systems were installed within isolation rooms, and used as a complementary disinfection procedure, being operated continuously during daylight hours in occupied rooms, under conditions where normal clinical care and infection control measures were implemented. The effect of the system was assessed through contact-plate sampling of bacterial levels on a range of frequently touched contact surfaces (e.g. locker top, bed table, bed rails, bin lids, light switches and door handles) which are commonly associated with being 'high-risk' surfaces for cross-transmission of HCAs, as well as surfaces likely to have high contamination levels due to aerial deposition, such as ledges. Samples were typically collected (i) before use, (ii) during use, and (iii) some time after the HINS-light EDS units had been switched off, with the same contact surfaces sampled throughout each study. Bacterial levels were assessed using 55 mm contact agar plates, with a surface area of 23.76 cm², which were inoculated by pressing the agar surface onto the environmental surface. Studies monitored the levels of staphylococcal bacteria (a good indicator of contamination of human origin), and the total viable bacteria levels in order to establish the effect of the system for reducing levels of bacterial contamination around the isolation room.^{58–60} For collection of staphylococcal organisms, Baird Parker with egg yolk telurite agar (BPA: a selective medium for the growth of staphylococcal-type organisms) contact plates were used. Tryptone soya agar (TSA) contact plates, which use non-selective growth medium, were used to obtain total viable bacterial counts (TVC). Microbiological assessment, as colony-forming unit counts, was based upon growth on the contact agar plates after incubation at 37°C for 24 h (TSA plates) or 48 h (BPA plates).

Several studies also characterized the staphylococcal isolates by subculturing selected isolates and then testing using Staphaurex Plus (Remel Europe Ltd, Dartford, UK) and PBP2 Latex Agglutination Test (Oxoid Ltd), to identify *S. aureus* and methicillin-resistant *S. aureus* isolates, respectively.

Inpatient studies

An initial study evaluated use of the system for disinfection of an unoccupied isolation room, and results demonstrated a significant 90% reduction ($P = 0.000$) in the staphylococcal contamination on surfaces around the room after 24 h use.⁵⁸ Studies in burns isolation rooms occupied by MRSA-positive patients, with treatment periods ranging from two to seven days, demonstrated that staphylococcal contamination on surfaces around the rooms was significantly reduced by 56–86%, over and above the reductions achieved by cleaning alone. Levels of presumptive *S. aureus* and MRSA showed similar reductions.⁵⁸ When use of the system ceased, recontamination of the room was observed, to levels similar to pre-treatment contamination levels.

An example of the data from one published study is shown in Figure 2, which demonstrates the mean reductions in the total staphylococcal counts and the presumptive *S. aureus* levels in an occupied burns unit isolation room, before, during and after five-day use of HINS-light EDS. Samples ($N = 70$) were collected twice during each of the three phases, and the results from all sampled surfaces have been pooled to demonstrate the overall decontamination effect the system had across the room. In this study, data demonstrated that a significant 62% decrease in total staphylococcal counts and 50% decrease in presumptive

S. aureus were achieved ($P < 0.05$) after five days' use of the system. 'After use' samples, collected during a six-day period after the system had been turned off, showed that contamination around the room had significantly risen, with 126% and 98% increases in the total staphylococci and presumptive *S. aureus* counts, respectively ($P < 0.05$), thus reinforcing the recontamination effect that occurs after removal of the light treatment.⁵⁸ Extended use of the system also proved to further reduce the bacterial contamination around the room, supporting the continuous use of this system for maintaining low contamination levels around isolation rooms.⁵⁸ Importantly, studies were performed to show that the decontamination effect was not patient or room dependent.⁵⁹

Studies carried out in an ICU isolation room also demonstrated system efficacy, with 60–70% reductions in both the staphylococcal and the total bacterial contamination across the entire sampled room environment.⁶⁰ In addition to demonstrating an overall reduction in contamination around the room, results demonstrated that exposed surfaces had reduced contamination levels as a result of use of the system, and an example of this is shown in Figure 3. Levels of bacteria on various surfaces around an occupied ICU isolation room were determined before use of the HINS-light EDS, and resampled after a five-day exposure period. Results demonstrated that despite marked variation in the initial bacterial bioburden there was a marked decrease in levels of bacterial contamination at all tested sites.

In addition to these findings, a significant factor noted in the studies carried out in the ICU isolation room was that despite asymmetrical positioning of the EDS units within the room, the special distribution of bacterial contamination was reduced almost uniformly across all the sampled contact surfaces. This suggested that disinfection of airborne bacteria contributes to the reductions in bacterial contamination levels, and the installation positions of the systems may not be critical.⁶⁰

Outpatient studies

In addition to its use for disinfection of occupied inpatient isolation rooms, the HINS-light EDS has also proved effective when used in an outpatient clinic.⁵⁹ Communal use of outpatient clinic rooms provides a recognized risk of cross-contamination between subsequently treated patients; therefore it is important to maintain cleanliness in these areas throughout the day. Studies were carried out to evaluate the environmental bacterial levels at the start and end of 8 h clinic sessions, with and without use of the EDS. A statistically significant 61% efficacy was achieved ($P = 0.02$), leading to the suggestion that use of this system would be beneficial in other similar communal patient rooms such as the bathroom or physiotherapy room, where decontamination of all surfaces is unachievable between each patient due to time limitations.⁵⁹

Overall, results have been successful, showing that use of 405 nm light achieves significant reductions in bacterial contamination levels around isolation room environments.^{58–60} Results also demonstrated that when switched off, the decontamination effect ceases and bacterial contamination levels return to around pre-treatment levels, further confirming the effectiveness of the 405 nm light. It is important to note that these results were achieved under a range of clinical conditions within a busy city hospital environment, and that the bacterial disinfection results obtained were over and above

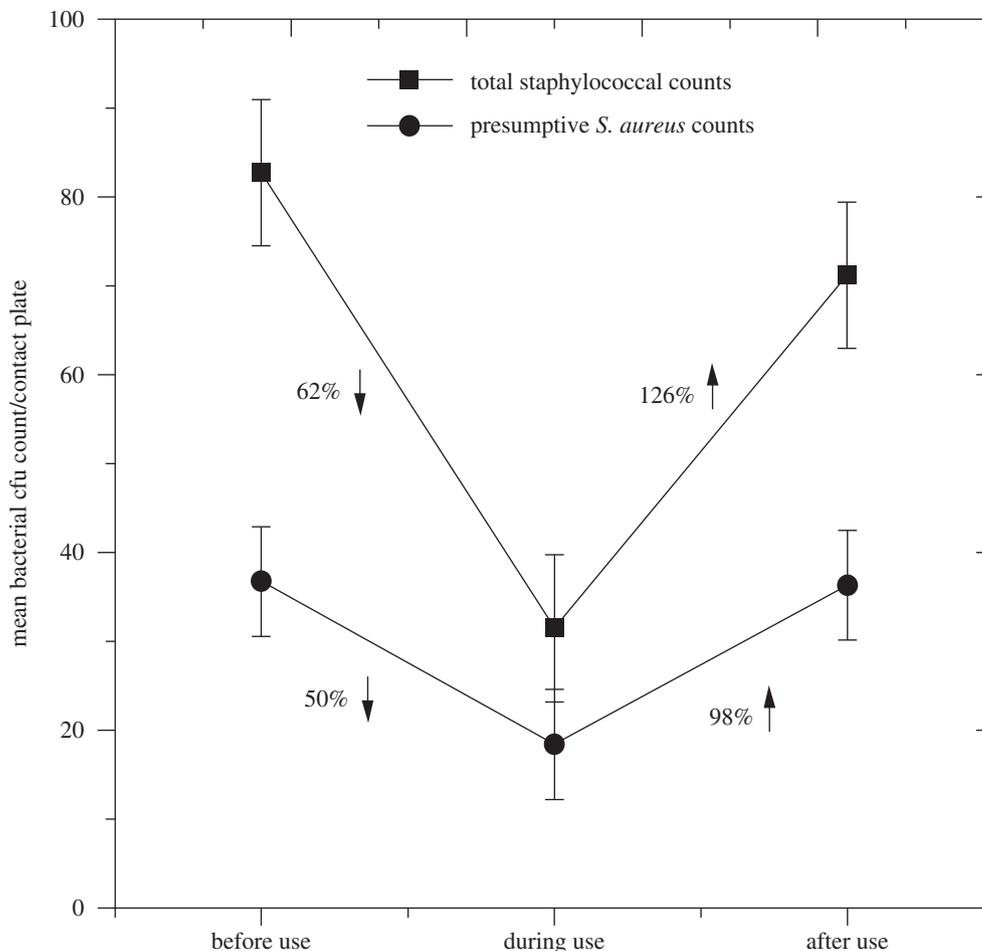


Figure 2. Mean reductions in the total staphylococcal colony-forming unit (cfu) counts and the presumptive *Staphylococcus aureus* levels across an occupied burns unit isolation room: before, during, and after five-day use of high-intensity narrow spectrum (HINS)-light environmental disinfection system (EDS). Contact plate samples ($N = 70$) were collected twice during each phase and the results pooled to assess the overall decontamination effect. A significant 62% decrease in total staphylococcal counts and a 50% decrease in presumptive *S. aureus* were achieved ($P < 0.05$). 'After use' samples showed that contamination around the room had significantly risen over the six days after the system had been switched off: 126% and 98% increase in the total staphylococci and presumptive *S. aureus* counts, respectively ($P < 0.05$). (Data adapted from Maclean et al.⁵⁸).

those achieved by the hospital's normal, stringent, infection control procedures which remained fully in place throughout the study.^{58–60} Further studies are required to establish the effectiveness of 405 nm light for disinfection of larger communal environments.

Comparison of 405 nm light with other environmental decontamination systems

Increased awareness of the importance of the hospital environment as a source of nosocomial pathogens has not only focused attention on improving the efficiency of conventional cleaning and disinfection procedures, but has also led to the development of a range of novel technologies for enhanced decontamination of whole-room environments, including new UV systems (as discussed earlier), steam cleaning, hydrogen peroxide vapour, and super-oxidized water fogging.^{7,65–67} Although these systems are effective for widespread disinfection of the room environment, they require, for safety reasons, experienced operator supervision and their use is restricted to

unoccupied, sealed rooms, thereby resulting in rooms being out-of-commission for periods of time – a consequence which can be costly and undesirable in busy ward areas. Additionally, whereas these systems provide effective decontamination, studies have found that once treatment has finished, there is rapid and widespread recontamination of the room.⁶⁸ In addition to human safety considerations, another problem associated with UV light and chemically based technologies is the potential for long-term material degradation of furniture and equipment within the treated room if these are repeatedly exposed.^{69,70} Therefore these methods are best-suited for terminal- and deep-cleaning procedures, but are ineffective for maintaining low levels of contamination.

Whereas UV irradiation and 405 nm light technology possess some similar features, they are in many respects quite distinct technologies both in their modes of action and methods of application (Table 1, Figure 1). Although UV light is strongly germicidal it is dangerous to humans, and the different UV waveband regions corresponding to UVC, UVB and UVA can cause a wide range of detrimental effects on the human eye and skin.⁷⁰ Violet–blue wavelengths within the visible

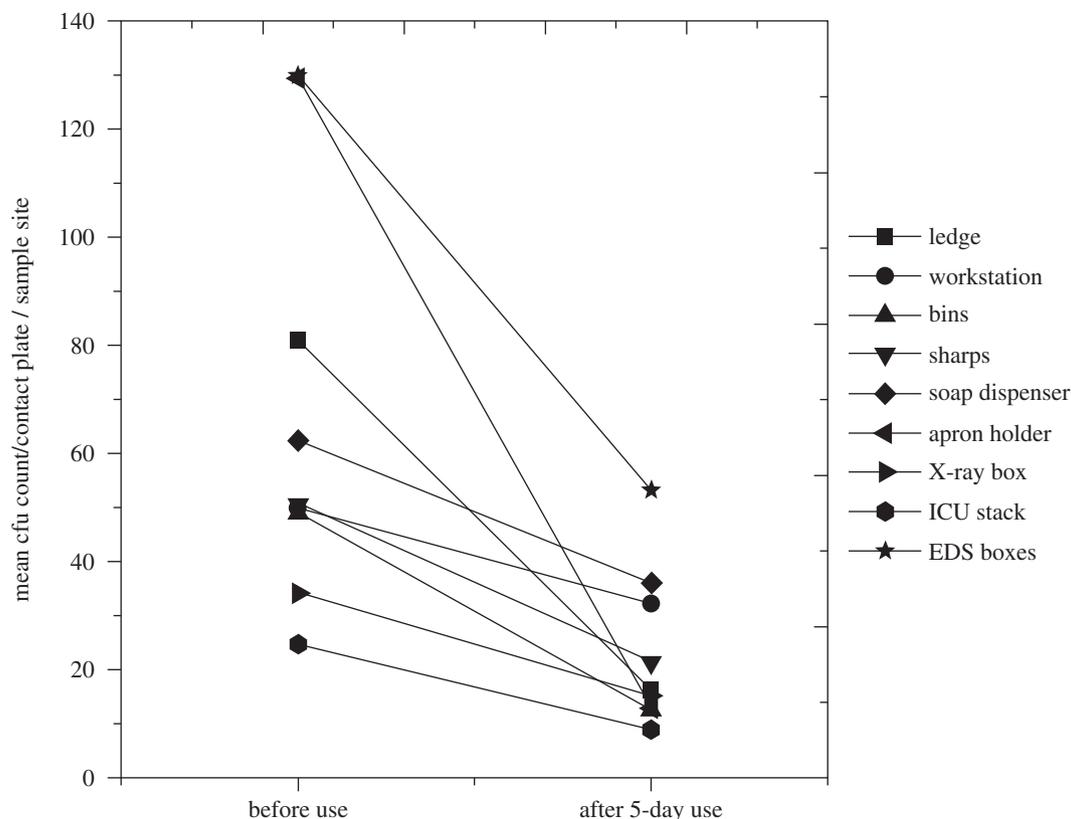


Figure 3. Reductions in the mean levels of environmental bacteria on a range of surfaces in an intensive care unit isolation room before and after five-day use of the high-intensity narrow spectrum (HINS)-light environmental disinfection system. Tryptone soya agar contact plate samples were collected from each surface and results pooled to show the mean reduction in contamination on the sampled surface. (Data adapted from Maclean *et al.*⁶⁰).

spectrum can also cause harmful effects at high irradiance levels, especially at 440 nm which can cause photoreinitis, and at 480 nm which is the peak sensitivity of mammalian photosensitive retinal ganglion cells (pRGCs) which modulate diverse physiological responses to light, including circadian physiology and pupil constriction.^{61,62,76} A comparison of the biological effects of radiation extending from the UV into the visible light regions is presented in Figure 1. Although 405 nm light is germicidal, it falls within a relatively benign wavelength region, and, if operated at appropriate irradiance levels, it is safe for human exposure.^{61,62}

The above features explain why the 405 nm light environmental disinfection technology, in comparison with other whole-room decontamination systems including UV technology, can be operated continuously in the presence of patients and staff, thus facilitating a background decontamination effect which maintains low levels of contamination.^{58–60} Continuous operation of the 405 nm light system ensures that there is a level of disinfection concurrently being applied even during periods of high activity, such as visiting hours, and bed and bandage changing.^{77,78} Whereas disinfectant cleaning and hand hygiene are critical for maintaining a clean environment and minimizing the spread of potential pathogens, compliance with handwashing tends to be low after direct contact with a patient, and healthcare workers are even less likely to wash their hands after being in contact with the environmental surfaces around the patients, even though these surfaces can be reservoirs of potential pathogens.⁷⁹ Use of the 405 nm light

technology can strategically augment this by enhancing the low levels of contamination achieved with intermittent cleaning, and also provide decontamination of surfaces within rooms, such as walls and high ledges, as well as delicate equipment, which may not be routinely cleaned using disinfectants. Moreover the system can be automatically operated with no user training required, and consequently problems with staff and patient compliance do not apply.^{58–60}

As with all methods of cleaning and disinfection, there are inherent disadvantages with any procedure. A limitation of the 405 nm light technology is that, to ensure that patient-friendly room illumination conditions are used, relatively low irradiance levels are applied and this impacts on microbial inactivation rates which are inevitably lower than can be achieved with other decontamination technologies albeit only in short-term comparisons. The high doses of 405 nm light required for inactivation of endospores means that it is unlikely that 405 nm light alone could be realistically applicable for the specific environmental decontamination of *C. difficile* spores. Nevertheless, enhancement of the inactivation may be achieved when combined with other decontamination methods such as oxidative biocides, due to the similar oxidative damage that is exerted on the bacteria by both treatments.³³ In addition to the resilience of spores, the antiviral efficacy of violet–blue light has not been fully established, and further research in this area is required. Also, similar to UVC technology, 405 nm light effectively treats hospital air, but only surfaces that are directly or reflectively exposed to the light are

Table 1
Comparison of the properties of ultraviolet C (UVC) and 405 nm light for environmental disinfection applications

	UVC light	405 nm light
Typical/potential use	Terminal clean of air- and light-exposed surfaces.	Continuous disinfection of air- and light-exposed surfaces.
Safety	Significant safety hazards associated with human exposure; can cause DNA mutations, erythema. ⁷⁰	Can be used safely in the presence of people at recommended irradiation levels. ^{58–60}
Mechanism of action	DNA damage kills cells. Sublethally damaged cells can recover using photoreactivation mechanism to repair DNA. ^{71,72}	Photo-excitation of intracellular molecules induces oxidation of microbial cells. No known repair mechanism. ^{73,74}
Antimicrobial activity	Broad-spectrum action against a range of micro-organisms including spores and viruses. ^{51,52}	Effective against bacteria, fungi, yeasts and spores; antiviral activity not yet fully established. ^{32,33,56}
Antimicrobial efficacy	Rapid inactivation rate within treatment zone. ^{6,24}	Comparably slower inactivation rate within treatment zone. ^{58–60}
Materials compatibility	UV-light-associated polymer damage. ⁶⁹	Lower energy 405 nm wavelengths more materials compatible. ⁶⁹
Ease of use for environmental disinfection	Rooms/wards need to be vacated during use; operator training required. ^{6,24}	Can be safely used during room occupation; no operator safety training required. ^{58–60}
Microbial mutagenic potential	Powerful mutagen that may encourage resistance development.	Multi-target oxidative action mitigates against resistance development. ⁷⁵
Penetrability	Does not penetrate through plastics and glass, and weakly penetrates into water and fabrics.	Can penetrate through plastics and glass, and penetrates into water and fabrics. ⁴⁴

treated, and the effects on occluded or darkly shadowed areas are limited. It is also the case that whereas all of the new technologies including 405 nm light can claim to have demonstrated enhanced disinfection of the hospital environment, **translation of this potential benefit into a significant reduction in infection rates will be required to ensure the widespread uptake of these new disinfection technologies.**

Further commentary regarding the application of 405 nm light for hospital disinfection

Regarding the deployment of the HINS-light system within hospitals, although important issues such as disinfection efficacy and patient safety have been addressed, other questions relating to the use of such a novel light source in clinical settings must also be considered. Undoubtedly enrichment of room lighting with additional violet–blue light will alter the normal lighting effect. This could have some impact on patient and staff comfort levels, and possible effects on medical procedures that involve colour perception must also be considered. In the hospital trials already conducted with the HINS-light EDS, no such issues have been problematic (unpublished observations) but monitoring for such effects must remain during uptake of this technology. Further hospital-based studies, funded by the Scottish Infection Research Network and the Chief Scientist Office, are currently being initiated to investigate the acceptability of the technology, and to ensure that the technology is optimized with staff and patient comfort fully taken into account. There may also be implications for colours employed in hospital furnishings and fabrics, as these may serve to amplify or suppress the reflection or absorption of violet–blue light.

As already discussed, a benefit of 405 nm light over UV-light for disinfection purposes is that, unlike UV-light, 405 nm light, because of its lower photon energy, **does not cause photo-**

degradation of photosensitive materials such as rubbers and plastics used in the hospital environment and equipment.⁶⁹

However, strong visible light can cause photochemical changes in light-sensitive solutions, and this aspect requires consideration if such solutions were to be exposed for long periods. At the relatively low 405 nm light intensities used, and considering the fact that light intensity decreases upon transmission through materials, e.g. plastic tubing or intravenous bag material, then this issue is not anticipated to be problematic but nevertheless must remain a consideration if highly light-sensitive pharmaceuticals were introduced.^{58–60}

The HINS-light system uses LED-based technology and, as such, it benefits from the well-established characteristics of LED lighting, namely reduced energy requirements, long operational (lifetime) use, and low maintenance characteristics. In the hospital trials already conducted, the HINS-light EDS unit is designed to be easily retrofitted into the ceiling in place of a ceiling tile. Installed units have remained maintenance-free and fully operational over the trial period, which now extends to several years. From a lighting technology perspective, it is interesting that the introduction of this LED-based disinfection system is concurrent with major potential changes taking place in general lighting technology. Considerable debate is underway regarding the advantages and disadvantages of replacing conventional fluorescent lighting with LED sources, a discussion that is mainly being driven by potential energy efficiency gains associated with LED lighting. Another potential advantage of LED technology is the capacity to blend different colours to 'fine tune' the colour spectrum to suit different environments and applications. In this context it is interesting that it is now appreciated, and as previously discussed in this review, that the nature of the light spectrum can affect circadian rhythmicity, sleep and mood and that this is associated with photosensitive retinal ganglion cells in the eye.⁷⁶ Such effects are important not only in the home and

workplace but also for patients in the hospital environment, where it has been suggested that more research is required to better understand how lighting in the hospital environment can influence sleep, mood and pain in medical inpatients.²² Future development of the HINS-light EDS system will undoubtedly be influenced by the various considerations outlined above.

Conclusions

Although the germicidal effects of sunlight and UV light have been known for more than a century, it is only comparatively recently that the antimicrobial properties of visible light in the violet–blue region of the spectrum have been recognized and studied in a number of laboratories. Given the severity of current and anticipated future microbiological problems faced by society, the development of any new antimicrobial weapon is to be welcomed. Violet–blue light, with particular efficacy at 405 nm, has been shown to possess broad-spectrum photodynamic antimicrobial activity, so its use has been suggested for a range of potential clinical and medical applications.

One such application is the use of 405 nm light for environmental disinfection. The increased safety of 405 nm light wavelengths compared to UV light has facilitated development of this light technology for safe, continuous disinfection of occupied environments, and results have shown the successful application of this system for environmental disinfection of hospital isolation rooms and clinics. This technology, termed the HINS-light EDS, has demonstrated a significant capability for reducing environmental bacterial contamination in clinical patient areas, over and above reductions achieved using the conventional cleaning and infection control strategies alone. In common with the aspirations of other novel, whole-room disinfection systems, it is intended that this intervention technology, when used in conjunction with conventional infection control procedures, may help reduce numbers of pathogens in the environment, thereby limiting the likelihood of pathogen transmission from the environment to patients, and thus contribute to reducing levels of HCAs.

Whereas violet–blue 405 nm light irradiation represents a new antimicrobial approach, the physical nature of this light source and the limitations of its antimicrobial effects must be understood. Inevitably microbial inactivation rates using 405 nm light are slower than can be achieved with the typical application of many other physical and chemical disinfection and sterilization treatments. This limitation is, however, mitigated by its operational facility for continuous application to disinfect air and all illuminated surfaces in occupied environments and by the biochemical mechanism of 405 nm light inactivation. The photodynamic inactivation process induced by 405 nm light exposure involves a multi-targeted intracellular killing effect resulting from the generation of ROS, a killing mechanism that is not conducive to microbial resistance development. Given these unique features, it is evident that 405 nm violet–blue light technology represents a novel antimicrobial approach that may make some contribution to tackling the challenge posed by ubiquitous environmental contamination, and to the ongoing health and resource problems associated with HCAs.

Conflict of interest statement

The intellectual property rights of the HINS-light EDS belong to the University of Strathclyde. The University has made all

systems for research purposes only and no commercial company manufactures this technology.

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None.

References

1. Stone PW. Economic burden of healthcare associated infections: an American perspective. *Expert Rev Pharmacoecon Outcomes Res* 2009;9:417–422.
2. Rutala WA, Weber DJ. Are room decontamination units needed to prevent transmission of environmental pathogens? *Infect Control Hosp Epidemiol* 2011;32:743–747.
3. Dancer SJ. Hospital cleaning in the 21st century. *Eur J Microbiol Infect Dis* 2011;12:1473–1481.
4. Sharma M, Hudson JB. Ozone gas is an effective and practical antibacterial agent. *Am J Infect Control* 2008;36:559–563.
5. Merandzic MN, Cadnum JL, Eckart KE, Donskey CJ. Evaluation of a handheld far-ultraviolet radiation device for decontamination of *Clostridium difficile* and other healthcare-associated pathogens. *BMC Infect Dis* 2012;12:120–125.
6. Otter JA, Yezli S, Perl TM, Barbut F, French GL. The role of ‘no-touch’ automated room disinfection systems in infection prevention control. *J Hosp Infect* 2013;83:1–13.
7. Andersen BM, Bånrud H, Bøe E, Bjordal O, Drangsholt F. Comparison of UV-C lights and chemicals for disinfection of surfaces in hospital isolation units. *Infect Control Hosp Epidemiol* 2006;27:729–734.
8. Nardell EA, Bucher SJ, Brickner PW, et al. Safety of upper room ultraviolet germicidal air disinfection for room occupants: results from the Tuberculosis Ultraviolet Shelter Study. *Public Health Rep* 2008;123:52–60.
9. Reed NG. The history of ultraviolet germicidal irradiation for air disinfection. *Public Health Rep* 2010;125:15–27.
10. Dai T, Gupta A, Huang YY, et al. Blue light eliminates community acquired methicillin resistant *Staphylococcus aureus* in infected mouse skin abrasions. *Photomed Laser Surg* 2013;31:531–538.
11. Dai T, Gupta A, Huang YY, et al. Blue light rescues mice from potentially fatal *Pseudomonas aeruginosa* burn infection: efficacy, safety and mechanism of action. *Antimicrob Agents Chemother* 2013;57:1238–1245.
12. Elman M, Slatkine M, Harth Y. The effective treatment of acne vulgaris by a high intensity narrow band 405–420 nm light source. *J Cosmet Laser Ther* 2003;5:111–116.
13. Shalita AR, Harth Y, Elman M, et al. Acne phototherapy using UV free high intensity narrow band blue light: a three centre clinical study. *Progress Biomed Optics Imaging* 2001;2:61–73.
14. Kleinpenning MM, Otero ME, van Erp PEJ, Gerritsen R, van de Kerkhof PCM. Efficacy of blue light versus red light in the treatment of psoriasis: a double blind, randomized comparative study. *J Eur Acad Dermatol Venereol* 2012;26:219–225.
15. Ganz RA, Viveiros J, Ahmad A, et al. *Helicobacter pylori* in patients can be killed by visible light. *Lasers Surg Med* 2005;36:260–265.
16. Lembo AJ, Ganz RA, Sheth S, et al. Treatment of *Helicobacter pylori* infection with intra-gastric violet light phototherapy: a pilot clinical trial. *Lasers Surg Med* 2009;41:337–344.
17. McDonald R, MacGregor SJ, Anderson JG, Maclean M, Grant MH. Effect of 405-nm high-intensity narrow-spectrum light on fibroblast populated collagen lattices – an in vitro model of wound healing. *J Biomed Optics* 2011;16:048003.
18. Kowalski W. *Ultraviolet germicidal irradiation handbook UVGI for air and surface disinfection*. Heidelberg: Springer; 2009.
19. Hobady RA. Sunlight therapy and solar architecture. *J Med Hist* 1997;4:455–472.
20. Hobday RA, Dancer SJ. Roles of sunlight and natural ventilation for controlling infection: historical and current perspectives. *J Hosp Infect* 2013;84:271–282.

21. Lieveer R, van Someren JW, Nielen MA, Uitdehaag BMJ, Smit JH, Hoogendijk WJG. Bright light treatment in elderly patients with non-seasonal major depressive disorder: a randomized placebo-controlled trial. *Am Med Assoc* 2011;**68**:61–70.
22. Bernhofer EI, Higgins PA, Daily BJ, Burant CJ, Hornick TR. Hospital lighting and its association with sleep, mood and pain in medical inpatients. *J Adv Nurs* 2014;**70**:1164–1173.
23. Kent A. News and views from the literature. *Rev Obstet Gynecol* 2013;**6**:25–38.
24. Simmons S, Morgan M, Hopkins T, Helsabeck K, Stachowiak J. Impact of a multi-hospital intervention utilising screening, hand hygiene education and pulsed xenon ultraviolet (PX-UV) on the rate of hospital associated methicillin *Staphylococcus aureus*. *J Infect Prev* 2013;**14**:172–174.
25. Boyce JM, Havill NL, Moore BA. Terminal decontamination of patient rooms using an automated mobile UV light unit. *Infect Control Hosp Epidemiol* 2011;**32**:737–742.
26. Davies A, Pottage T, Bennett A, Walker J. Gaseous and air decontamination technologies for *Clostridium difficile* in the healthcare environment. *J Hosp Infect* 2011;**77**:199–203.
27. Shin GA, Linden KG, Arrowood MJ, Sobsey MD. Low pressure UV inactivation and DNA repair potential of *Cryptosporidium parvum* oocysts. *Appl Environ Microbiol* 2001;**67**:3029–3032.
28. Kac G, Gueneret M, Rodi A, et al. Evaluation of new disinfection procedure for ultrasound probes using ultraviolet light. *J Hosp Infect* 2007;**65**:163–168.
29. Hamblin MR, Viveiros J, Yang C, Ahmadi A, Ganz RA, Tolckoff MJ. *Helicobacter pylori* accumulates photoactive porphyrins and is killed by visible light. *Antimicrob Agents Chemother* 2005;**49**:2822–2827.
30. Guffey JS, Wilborn J. *In vitro* bactericidal effects of 405-nm and 470-nm blue light. *Photomed Laser Surg* 2006;**24**:684–688.
31. Enwemeka CS, Williams D, Hollosi S, Yens D, Enwemeka SK. Visible 405nm SLD photo destroys methicillin-resistant *Staphylococcus aureus* (MRSA) *in vitro*. *Laser Surg Med* 2008;**40**:734–737.
32. Maclean M, MacGregor SJ, Anderson JG, Woolsey GA. Inactivation of bacterial pathogens following exposure to light from a 405nm LED array. *Appl Environ Microbiol* 2009;**75**:1932–1937.
33. Maclean M, Murdoch LE, MacGregor SJ, Anderson JG. Sporicidal effects of high-intensity 405 nm visible light on endospore-forming bacteria. *Photochem Photobiol* 2013;**89**:120–126.
34. Endarko E, Maclean M, Timoshkin IV, MacGregor SJ, Anderson JG. High intensity 405 nm light inactivation of *Listeria monocytogenes*. *Photochem Photobiol* 2012;**88**:1280–1286.
35. Maclean M, MacGregor SJ, Anderson JG, Woolsey GA. High-intensity narrow-spectrum light inactivation and wavelength sensitivity of *Staphylococcus aureus*. *FEMS Microbiol Lett* 2008;**285**:227–232.
36. Goldoni A, Porphyrins: fascinating molecules with biological significance. *ELETTA Laboratory, research highlights 2001–2002: atomic, molecular and supramolecular studies* 2002. p. 64–65.
37. Ashkenazi H, Malik Z, Harth Y, Nitzan Y. Eradication of *Propionibacterium acnes* by its endogenous porphyrins after illumination with high intensity blue light. *FEMS Immunol Med Microbiol* 2003;**35**:17–24.
38. Feuerstein O, Ginsburg I, Dayan E, Veler D, Weiss EI. Mechanism of visible light phototoxicity on *Porphyromonas gingivalis* and *Fusobacterium nucleatum*. *Photochem Photobiol* 2005;**81**:1186–1189.
39. Maclean M, MacGregor SJ, Anderson JG, Woolsey GA. The role of oxygen in the visible-light inactivation of *Staphylococcus aureus*. *J Photochem Photobiol B* 2008;**92**:180–184.
40. Lipovsky A, Nitzan Y, Friedmann H, Lubart R. Sensitivity of *Staphylococcus aureus* strains to broadband visible light. *Photochem Photobiol* 2009;**85**:255–260.
41. Dai T, Gupta A, Murray CK, Vrahas MS, Tegos GP, Hamblin MR. Blue light for infectious diseases: *Propionibacterium acnes*, *Helicobacter pylori*, and beyond? *Drug Resist Update* 2012;**15**:223–236.
42. Murdoch LE, Maclean M, Endarko E, MacGregor SJ, Anderson JG. Bactericidal effects of 405-nm light exposure demonstrated by inactivation of *Escherichia*, *Salmonella*, *Shigella*, *Listeria* and *Mycobacterium* species in liquid suspensions and on exposed surfaces. *Scient World J* 2012;**2012**:137805.
43. Wasson CJ, Zourelis JL, Aardsma NA, et al. Inhibitory effects of 405 nm irradiation on *Chlamydia trachomatis* growth and characterization of the ensuing inflammatory response in HeLa cells. *BMC Microbiol* 2012;**12**:176–186.
44. McKenzie K, Maclean M, Timoshkin IV, Endarko E, MacGregor SJ, Anderson JG. Photoinactivation of bacteria attached to glass and acrylic surfaces by 405 nm light: potential application for biofilm decontamination. *Photochem Photobiol* 2013;**89**:927–935.
45. Wilcox MH, Fawley WN. Hospital disinfectants and spore formation by *Clostridium difficile*. *Lancet* 2000;**356**:1324.
46. Rupnik M, Wilcox MH, Gerding DN. *Clostridium difficile* infection: new developments in epidemiology and pathogenesis. *Nature* 2009;**7**:526–536.
47. St Denis TG, Dai T, Izikson L, et al. All you need is light antimicrobial photo inactivation as an evolving and emerging discovery strategy against infectious disease. *Landes Biosci* 2011;**2**:509–520.
48. McKenzie K, Maclean M, Timoshkin IV, MacGregor SJ, Anderson JG. Bactericidal effect of 405nm light on *Escherichia coli* and *Listeria monocytogenes* in the presence of sub-lethal stress. *Int J Food Microbiol* 2014;**170**:91–98.
49. Donlan MR. Biofilms: microbial life on surfaces. *Emerg Infect Dis* 2002;**8**:881–890.
50. Hota S, Hirji Z, Stockton K, et al. Outbreak of multidrug resistant *Pseudomonas aeruginosa* colonization and infection secondary to imperfect intensive care unit room design. *Infect Control Hosp Epidemiol* 2009;**30**:25–33.
51. Chang JCH, Ossoff SF, Lobe DC, et al. UV inactivation of pathogenic and indicator microorganisms. *Appl Environ Microbiol* 1985;**49**:1361–1365.
52. Hijnen WAM, Beerendonk EF, Medema GJ. Inactivation credit of UV radiation for viruses, bacteria and protozoan (oo)cysts in water: a review. *Water Res* 2006;**40**:3–22.
53. Murdoch LE, Maclean M, MacGregor SJ, Anderson JG. Inactivation of *Campylobacter jejuni* by exposure to high-intensity 405-nm visible light. *Foodborne Path Dis* 2010;**7**:1211–1216.
54. Feuerstein O, Persman N, Weiss EI. Phototoxic effect of visible light on *Porphyromonas gingivalis* and *Fusobacterium nucleatum*: an *in vitro* study. *Photochem Photobiol* 2004;**80**:412–415.
55. Soukos NS, Som S, Abernethy AD, et al. Phototargeting oral black-pigmented bacteria. *Antimicrob Agents Chemother* 2005;**49**:1391–1396.
56. Murdoch LE, McKenzie K, Maclean M, MacGregor SJ, Anderson JG. Lethal effects of high intensity violet 405-nm light on *Saccharomyces cerevisiae*, *Candida albicans* and on dormant and germinating spores of *Aspergillus niger*. *Fungal Biol* 2013;**117**:519–527.
57. Yin R, Dai T, Avci P, et al. Light based anti-infectives: ultraviolet C irradiation, photodynamic therapy, blue light and beyond. *Curr Opin Pharmacol* 2013;**13**:1–32.
58. Maclean M, MacGregor SJ, Anderson JG, et al. Environmental decontamination of a hospital isolation room using high-intensity narrow-spectrum light. *J Hosp Infect* 2010;**76**:247–251.
59. Bache SE, Maclean M, MacGregor SJ, et al. Clinical studies of the HINS-light environmental decontamination system for continuous disinfection in the burn unit inpatient and outpatient settings. *Burns* 2012;**38**:69–76.
60. Maclean M, Booth M, MacGregor SJ, et al. Continuous decontamination of an intensive care isolation room during patient occupancy using 405 nm light technology. *J Infect Prevent* 2013;**14**:176–181.
61. International Commission of Non-Ionizing Radiation Protection (ICNIRP). Guidelines on limits of exposure to optical radiation from 0.38–3.9 mm. *Health Physics* 1997;**73**:539–554.
62. International Commission on Non-Ionizing Radiation Protection (ICNIRP). Guidelines on limits of exposure to ultraviolet radiation of wavelengths between 180 nm and 400 nm (incoherent radiation). *Health Physics* 2004;**87**:171–186.

63. McDonald RS, Gupta S, Maclean M, et al. 405 nm light exposure of osteoblasts and inactivation of bacterial isolates from arthroplasty patients: potential for new decontamination applications? *Eur Cell Mater* 2013;25:204–214.
64. Bache SE, Maclean M, Anderson JG, et al. Laboratory inactivation of healthcare-associated isolates by a visible HINS-light source and its clinical application in the burns unit (Abstract). *Burns* 2011;37:S6.
65. French GL, Otter JA, Shannon KP, Adams NMT, Watling D, Parks MJ. Tackling contamination of the hospital environment by methicillin resistant *Staphylococcus aureus* (MRSA): a comparison between conventional terminal cleaning and hydrogen peroxide vapour decontamination. *J Hosp Infect* 2004;57:31–37.
66. Clark J, Barrett SP, Rogers M, Stapelton R. Efficacy of super oxidised water fogging in environmental decontamination. *J Hosp Infect* 2006;64:386–390.
67. Department of Health. *An integrated approach to hospital cleaning: microfiber cloth and steam cleaning technology*. London: DH; 2007.
68. Hardy KJ, Gossain S, Henderson N. Rapid recontamination with MRSA of the environment of an intensive care unit after decontamination with hydrogen peroxide vapour. *J Hosp Infect* 2007;66:360–368.
69. Andrady AL, Hamid SH, Hu X, Torikai A. Effects of increased solar ultraviolet radiation on materials. *J Photochem Photobiol B* 1998;46:96–103.
70. Matsumura Y, Ananthaswamy HN. Toxic effects of ultraviolet radiation on the skin. *Toxicol Appl Pharmacol* 2004;195:298–308.
71. Sinah RP, Hader DP. UV-induced DNA damage and repair: a review. *Photochem Photobiol Sci* 2002;1:225–236.
72. Oguma K, Katayama H, Ohgaki S. Photoreactivation of *Escherichia coli* after low or medium pressure UV disinfection determined by an endonuclease sensitive site assay. *Appl Environ Microbiol* 2002;68:6029–6035.
73. Hamblin MR, Hasan T. Photodynamic therapy: a new antimicrobial approach to infectious disease? *Photochem Photobiol Sci* 2004;3:436–450.
74. Nitzan Y, Kauffman M. Endogenous porphyrin production in bacteria by δ -aminolaevulinic acid and subsequent bacterial photo-eradication. *Laser Med Sci* 1999;14:269–277.
75. Donnelly RF, McCarron PA, Tunney MM. Antifungal photodynamic therapy. *Microbiol Res* 2008;163:1–12.
76. Foster RG. The 'third' photoreceptor system of the eye – photosensitive retinal ganglion cells. *Eur Ophthalmol Rev* 2009;2:84–86.
77. Shiomori T, Miyamoto H, Makishima K, et al. Evaluation of bedmaking-related airborne and surface methicillin-resistant *Staphylococcus aureus* contamination. *J Hosp Infect* 2002;50:30–35.
78. Bache SE, Maclean M, Gettinby G, Anderson JG, MacGregor SJ, Taggart I. Quantifying bacterial transfer from patients to staff during burns dressing and bed changes: implications for infection control. *Burns* 2013;39:220–228.
79. Bhalla A, Pultz NJ, Gries DM, et al. Acquisition of nosocomial pathogens on hands after contact with environmental surfaces near hospitalized patients. *Infect Control Epidemiol* 2004;25:164–166.